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Nuclear and Mitochondrial Gene Data Support Recent Radiation within the Sea Spider Species Complex *Pallenopsis patagonica*

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The climate history of the Antarctic continental shelf has formed a diverse benthic ecosystem over evolutionary time scales. The extent of faunal diversity has only recently been unveiled especially by using genetic data. In addition to newly reported species, known species of benthic invertebrates in the Southern Ocean turned out to be in fact species complexes representing genetically very distinct clades. Previous studies have shown that the sea spider *Pallenopsis patagonica* is such a species complex consisting of several divergent mitochondrial clades. However, genetic analyses of another sea spider complex, *Colossendeis megalonyx*, showed that looking at one mitochondrial gene only can lead to overestimation of species number within a species complex and revealed mito-nuclear discordances. In this study we expand the current data set of *P. patagonica* by adding not only samples from Patagonia, the Subantarctic and the Eastern Weddell Sea, but also sequence data for the nuclear internal transcribed spacer (ITS) region to obtain more information about the species complex. In fact, the number of distinct clades is reduced when looking at nuclear data, but there are no cases of mito-nuclear discordance and hence no evidence for hybridization and speciation reversal events between divergent mitochondrial clades as in *C. megalonyx*. As patterns of mitochondrial COI diversity and divergence within *P. patagonica* and *C. megalonyx* are very similar and molecular dating analyses of both species complexes suggest a recent separation of clades during the Pleistocene, different biological processes seem to have led to fast and stable species boundaries in *P. patagonica* as opposed to *C. megalonyx* where hybridization even across major mitochondrial lineages occurred.

Keywords: Southern Ocean, Chelicerata, Pycnogonida, DNA-barcoding, ITS, cryptic species

INTRODUCTION

Increased sampling of Southern Ocean habitats and the application of molecular taxonomy uncovered that Antarctic biodiversity has been drastically underestimated (Gutt et al., 2004; De Broyer and Danis, 2011; De Broyer et al., 2014). The main reason for the significant boost in number of newly found species is the detection of morphologically cryptic species in basically all studied taxonomic groups (see Janosik and Halanych, 2010; Kaiser et al., 2013 for reviews). Therefore, paradoxically the Southern Ocean has emerged from being regarded as a biodiversity sink to a center of marine biodiversity in the past two decades. This phenomenon of high *in situ* species diversity has been termed the Antarctic diversity pump (sensu Clarke and Crame, 1989).

Several processes have been discussed as drivers fueling this diversity pump (Clarke and Crame, 1989). In this context, molecular data have led to a paradigm shift: Most of the cryptic species have rendered the distribution ranges of formerly described species from broad (i.e., circum-Antarctic) to small and allopatric (Lörz et al., 2009; Held, 2014). As the timing of many of these divergence events was rather recent in the Plio- or Pleistocene (see Convey et al., 2009 for a review), an influence of the recurrent large-scale glaciations in these periods has been suggested as a main driver fueling the diversity pump, mainly through random genetic drift and lineage sorting in independent glacial refugia (Thatje et al., 2005, 2008; Allcock and Strugnell, 2012). Prominent signatures of population bottlenecks, in particular for shallow-water organisms, have supported that view (e.g., Janko et al., 2007; Raupach et al., 2010).

In this context, Pycnogonida or sea spiders have attracted particular attention, since they show an exceptionally high species diversity in the Southern Ocean (Clarke and Johnston, 2003; Munilla and Soler Membrives, 2009; Griffiths et al., 2011). Moreover, many pycnogonids are benthic brooders with probably limited dispersal capacity. Therefore, lineage sorting events in glacial refugia driving speciation should have been stronger than in other, free-spawning taxa (Allcock and Strugnell, 2012).

In agreement with these predictions, several molecular studies have reported evidence for overlooked diversity in various sea spider species (Mahon et al., 2008; Krabbe et al., 2010; Dietz et al., 2015a,b). Furthermore, intraspecific diversity was found to be significantly partitioned regionally, indicating limited gene flow (Arango et al., 2011; Dietz et al., 2015b; Dömel et al., 2015). Two species complexes stand out by far in terms of their identified numbers of mitochondrial clades revealed by classical DNA barcoding (amplification of cytochrome c oxidase subunit I gene, COI): *Colossendeis megalonyx* (Hoek, 1881) (Krabbe et al., 2010; Dietz et al., 2015b) and *Pallenopsis patagonica* (Hoek, 1881) (Weis et al., 2014; Harder et al., 2016). For *C. megalonyx*, Krabbe et al. (2010) reported the presence of six distinct mitochondrial lineages that likely represent cryptic species with mostly small and allopatric distribution ranges. However, extending the sampling range substantially Dietz et al. (2015b) revealed a much greater number of mitochondrial clades. These also showed mostly circum-Antarctic distribution

instead of regional partitioning. Similar findings were made for the crinoid *Promachocrinus kerguelensis* [Wilson et al. (2007): restricted distribution range; Hemery et al. (2012): circumpolar distribution range with extended data set]. Interestingly, analyses of a nuclear gene of *C. megalonyx* indicated that several of the mitochondrial clades do not represent distinct species as they had identical sequences for the otherwise highly variable nuclear internal transcribed spacer region (ITS; Dietz et al., 2015b). This suggests hybridization events between several mitochondrial clades and subsequently speciation reversal after the completion of mitochondrial lineage sorting within *C. megalonyx*.

For *P. patagonica*, Weis et al. (2014) showed that it is also a species complex as has been anticipated by Gordon (1944) and Pushkin (1975, 1993), and described a new species (*P. yepayekae* Weis, 2014 in Weis et al., 2014) using integrative taxonomy combining mitochondrial sequences with morphological characters. Harder et al. (2016) found evidence for even more diversity within the complex by adding further mitochondrial data specifically for Antarctic specimens. As Dietz et al. (2015b) have shown, only looking at mitochondrial data can lead to an overestimation of species number. Therefore nuclear data are needed for *P. patagonica* to explicitly test whether the identified mitochondrial clades reported by Weis et al. (2014) and Harder et al. (2016) are supported by such independent markers. Thus, in this study we analyzed both mitochondrial and nuclear data of the *P. patagonica* complex for a substantially extended data set as compared to Weis et al. (2014) and Harder et al. (2016). We hypothesized that (i) extending the data set for *P. patagonica* by new samples, specifically from previously unsampled locations, reveals further distinct mitochondrial clades, (ii) the number of distinct species is substantially smaller than the number of mitochondrial lineages when analyzing an independent nuclear gene marker, and (iii) the extended data set reveals broader distribution ranges for previously reported clades. Moreover, we addressed the significance of our results in the context of currently discussed evolutionary mechanisms generating Southern Ocean benthic diversity.

MATERIALS AND METHODS

Specimens and Sampling Sites

For the remainder of this study, we use the term *P. patagonica* sensu lato (s.l.) when referring to the whole species complex including *P. yepayekae*, because it groups within clades morphologically originally identified as *P. patagonica*. Individuals of *P. patagonica* s.l. from the shelf of South America, Subantarctic islands as well as around the Antarctic continent were analyzed (Table 1, Figure 1). Chilean specimens were collected by divers during Huinay Fjordos expeditions (HF16, HF21, HF24, and HF26). Falkland samples (ZDLT1) were provided by Vladimir Laptikhovskiy (Falkland Islands Fisheries Department, Stanley, Falkland Islands). Samples from the Southern Ocean were collected using different bottom trawls during several cruises on board the RRS *James Clark Ross* (British Antarctic Survey, Cambridge, UK) and the RV *Polarstern* (Alfred Wegener Institute Helmholtz

TABLE 1 | Specimens list for *P. patagonica* s.l. and outgroups used during study. Species names are only given for outgroups and *P. yepayekae* as other morphological determinations are as yet not possible. Sampling details (location, latitude, longitude, depth) and haplotype information for the specimens analyzed (molecular clade, sequence availability).

Species	Clade	Name	Lat	Lon	Depth	ZSM-Voucher Number	COI-GenBank/ BOLD Number	ITS-GenBank/ BOLD Number
	ANT_A	KT982317	-68.020	-67.671	208		KT982317	KY272398
	ANT_A	KT982356	-76.479	-165.738	457		KT982356	KY272399
	ANT_B	FJ969369	-71.621	-170.867	205		FJ969369	
	ANT_C	KT982322	-64.035	-56.728	220		KT982322	KY272414
	ANT_C	KT982333	-63.686	-56.859	400		KT982333	KY272415
	ANT_C	KT982334	-63.686	-56.859	400		KT982334	KY272416
	ANT_C	KT982341	-63.754	-55.684	334		KT982341	KY272417
	ANT_C	KT982343	-63.754	-55.684	334		KT982343	KY272418
	ANT_C	PpaA_001	-71.136	-11.527	123		KC794958	
	ANT_C	PS82_121_1	-76.966	-32.945		ZSM-A20160626	KY272315	KY272412
	ANT_C	PS82_143_2_1	-76.967	-32.866		ZSM-A20160623	KY272311	KY272419
	ANT_C	PS82_143_2_3	-76.967	-32.866		ZSM-A20160625	KY272319	KY272405
	ANT_C	PS82_156_2_1	-75.507	-27.486		ZSM-A20160629	KY272313	KY272407
	ANT_C	PS82_156_2_2	-75.507	-27.486		ZSM-A20160630	KY272309	KY272413
	ANT_C	PS82_156_2_3	-75.507	-27.486		ZSM-A20160631	KY272310	KY272410
	ANT_C	PS82_170_1	-74.906	-26.685		ZSM-A20160632	KY272318	KY272411
	ANT_C	PS82_170_2	-74.906	-26.685		ZSM-A20160633	KY272317	KY272409
	ANT_C	PS82_174_3	-74.491	-30.977		ZSM-A20160637	KY272312	KY272403
	ANT_C	PS82_223_1	-75.522	-28.973		ZSM-A20160730	KY272308	KY272408
	ANT_C	PS82_25_2_1	-74.705	-29.900		ZSM-A20160635	KY272314	KY272404
	ANT_C	PS82_25_2_2	-74.705	-29.900		ZSM-A20160636	KY272316	KY272406
	ANT_D.1	KT982325	-63.576	-54.629	227		KT982325	KY272396
	ANT_D.1	KT982326	-62.442	-55.459	245		KT982326	
	ANT_D.1	KT982330	-63.389	-60.120	310		KT982330	
	ANT_D.1	KT982331	-63.389	-60.120	310		KT982331	
	ANT_D.1	KT982346	-63.834	-62.664	256		KT982346	KY272397
	ANT_D.2	JR262_1058	-55.144	-36.245	195.21	ZSM-A20160708	KY272301	
	ANT_D.2	JR262_1319	-55.002	-37.272	148.81	ZSM-A20160709	KY272302	
	ANT_D.2	JR262_1597_2	-54.396	-37.384	174.98	ZSM-A20160710	KY272305	
	ANT_D.2	JR262_1903_1	-53.597	-41.214	132.83	ZSM-A20160711	KY272303	
	ANT_D.2	JR262_48_5_1	-54.284	-36.083	124.08	ZSM-A20160712	KY272297	
	ANT_D.2	JR262_48_5_2	-54.284	-36.083	124.08	ZSM-A20160713	KY272298	
	ANT_D.2	JR262_702_1	-55.166	-35.485	126.99	ZSM-A20160714	KY272299	
	ANT_D.2	JR262_744	-55.167	-35.485	126.84	ZSM-A20160715	KY272304	
	ANT_D.2	JR262_806_4	-54.984	-35.762	139.38	ZSM-A20160716	KY272300	
	ANT_D.2	JR287_124_1	-53.764	-36.681	151	ZSM-A20160691	KY272295	KY272393
	ANT_D.2	JR287_124_2	-53.764	-36.681	151	ZSM-A20160692	KY272294	KY272391
	ANT_D.2	JR287_124_3	-53.764	-36.681	151	ZSM-A20160693	KY272296	KY272394
	ANT_D.2	JR287_152	-53.758	-36.690	145	ZSM-A20160694	KY272292	
	ANT_D.2	JR287_191	-53.751	-36.699	145	ZSM-A20160695	KY272307	
	ANT_D.2	JR287_59_2	-54.944	-35.979	246	ZSM-A20160687	KY272293	KY272392
	ANT_D.2	PpaE_001_HT26	-53.461	-41.261	193	ZSM-A20160717	KC794959	
	ANT_D.2	PS77_211_6_1_3	-53.402	-42.668	290.2	ZSM-A20160696	KY272306	KY272395
	ANT_E	KT982297	-72.177	-103.514	341		KT982297	KY272442
	ANT_E	KT982318	-68.020	-67.671	208		KT982318	
	ANT_F	HM426218	-71.092	-11.508			HM426218	
	ANT_F	KT982324	-63.686	-56.859	400		KT982324	KY272429
	ANT_F	KT982332	-64.134	-56.860	310		KT982332	
	ANT_F	KT982342	-63.754	-55.684	334		KT982342	

(Continued)

TABLE 1 | Continued

Species	Clade	Name	Lat	Lon	Depth	ZSM-Voucher Number	COI-GenBank/ BOLD Number	ITS-GenBank/ BOLD Number
	ANT_F	PS77_226_7_1_1	-64.915	-60.621	226.2	ZSM-A20160648	KY272331	KY272434
	ANT_F	PS77_226_7_1_2	-64.915	-60.621	226.2	ZSM-A20160649	KY272334	KY272430
	ANT_F	PS77_248_2_2	-65.955	-60.466	212	ZSM-A20160643	KY272335	KY272433
	ANT_F	PS77_248_3_2_1	-65.924	-60.332	433	ZSM-A20160644	KY272337	KY272436
	ANT_F	PS77_248_3_2_2	-65.924	-60.332	433	ZSM-A20160645	KY272336	KY272431
	ANT_F	PS77_248_3_2_3	-65.924	-60.332	433	ZSM-A20160646	KY272338	KY272435
	ANT_F	PS77_248_3_2_4	-65.924	-60.332	433	ZSM-A20160647	KY272332	
	ANT_F	PS77_257_2_2_3	-64.913	-60.648	152.5	ZSM-A20160650	KY272330	KY272440
	ANT_F	PS77_257_2_2_5	-64.913	-60.648	152.5	ZSM-A20160651	KY272329	KY272432
	ANT_F	PS77_275	-70.940	-10.489		ZSM-A20160728	KY272326	KY272439
	ANT_F	PS77_291_1_2	-70.842	-10.587	267.5	ZSM-A20160642	KY272333	KY272437
	ANT_F	PS77_292_2_5	-70.846	-10.593	243.5	ZSM-A20160729	KY272327	KY272441
	ANT_F	PS82_58_1	-76.322	-28.992		ZSM-A20160627	KY272328	KY272438
	ANT_G	FJ969367	-71.258	-170.635	466		FJ969367	
	ANT_G	FJ969368	-72.014	-170.775	236		FJ969368	
	ANT_H	KT982338	-63.754	-55.684	334		KT982338	KY272422
	ANT_H	KT982352	-64.411	-61.963	664		KT982352	
	ANT_H	KT982354	-64.411	-61.963	664		KT982354	
	ANT_I	KT982316	-62.933	-61.479	188		KT982316	KY272423
	ANT_J	KT982293	-76.998	-175.093	541		KT982293	
	ANT_J	KT982294	-76.904	169.965	764		KT982294	KY272427
	ANT_J	KT982306	-76.998	-175.093	541		KT982306	KY272426
	ANT_J	KT982313	-76.998	-175.093	541		KT982313	KY272428
	ANT_K	PS82_143_2_2	-76.967	-32.866		ZSM-A20160624	KY272325	KY272425
	ANT_K	PS82_244_4	-72.799	-19.495		ZSM-A20160640	KY272323	
	ANT_K	PS82_246_2	-70.928	-10.475		ZSM-A20160641	KY272324	KY272424
	ANT_L	PS82_109_2_2	-77.016	-33.695		ZSM-A20160622	KY272339	KY272420
	ANT_L	PS82_34_2	-76.069	-30.160		ZSM-A20160628	KY272340	KY272421
	ANT_M	HM426171	-71.317	-13.942			HM426171	
	ANT_M	PS82_183_1_1	-74.250	-37.749		ZSM-A20160638	KY272321	KY272400
	ANT_M	PS82_183_1_2	-74.250	-37.749		ZSM-A20160639	KY272320	KY272401
	ANT_M	PS82_240_2	-74.660	-28.763		ZSM-A20160731	KY272322	KY272402
	ANT_N	PpaE_002_HT25	-54.016	-37.437	78	ZSM-A20160718	KC794960	
	ANT_N	PS77_211_6_1_4	-53.402	-42.668	290.2	ZSM-A20160697	KY272360	KY272458
	SUB_1	PS77_208_5_1_1	-56.168	-54.548	292	ZSM-A20160726	KY272289	KY272367
	SUB_1	PS77_208_5_1_4	-56.168	-54.548	292	ZSM-A20160689	KY272288	
	SUB_2	ZSMA20111352_HT27	-51.269	-62.952	171-174	ZSM-A20111352	KF603937/CFAP037-11	
	SUB_2.1	HF26_254	-53.007	-73.923	31	ZSM-A20160456	KY272290	KY272368
	SUB_2.2	PS77_208_3	-56.152	-54.530	285.5	ZSM-A20160725	KY272291	KY272366
	SUB_3	ZSMA20111008_HT28	-50.414	-74.559	15-20	ZSM-A20111008	KF603952/CFAP026-11	KY272390
	SUB_4	PpaE_004_HT18	-52.574	-60.084	378	ZSM-A20160719	KC794961	KY272443
	SUB_4	PpaE_005_HT15	-52.574	-60.084	378	ZSM-A20160720	KC794962	
	SUB_4	PpaE_006_HT17	-52.574	-60.084	378	ZSM-A20160721	KC794963	KY272457
	SUB_4	PpaE_007_HT15	-52.574	-60.084	378	ZSM-A20160722	KC794964	
	SUB_4	PpaE_008_HT15	-52.574	-60.084	378	ZSM-A20160723	KC794965	
	SUB_4	PpaE_010_HT15	-52.962	-60.143	378	ZSM-A20160724	KC794966	
	SUB_4	PS77_208_5_1_2	-56.168	-54.548	292	ZSM-A20160727	KY272356	
	SUB_4	ZDLT1_889_1	-50.252	-61.567	159	ZSM-A20160698	KY272357	KY272445
	SUB_4	ZDLT1_889_2	-50.252	-61.567	159	ZSM-A20160699	KY272358	KY272446

(Continued)

TABLE 1 | Continued

Species	Clade	Name	Lat	Lon	Depth	ZSM-Voucher Number	COI-GenBank/ BOLD Number	ITS-GenBank/ BOLD Number
	SUB_4	ZDLT1_889_3	-50.252	-61.567	159	ZSM-A20160700	KY272359	KY272444
	SUB_4	ZSMA20111348_HT14	-50.434	-62.768	146-148	ZSM-A20111348	KF603953/CFAP027-11	
	SUB_4	ZSMA20111349_HT13	-51.269	-62.952	171-174	ZSM-A20111349	KF603960/CFAP034-11	
	SUB_4	ZSMA20111350_HT15	-51.269	-62.952	171-174	ZSM-A20111350	KF603961/CFAP035-11	
	SUB_4	ZSMA20111351_HT20	-51.269	-62.952	171-174	ZSM-A20111351	KF603962/CFAP036-11	
	SUB_4	ZSMA20111354_HT17	-51.086	-61.733	174-176	ZSM-A20111354	KF603954/CFAP028-11	
	SUB_4	ZSMA20111355_HT18	-51.086	-61.733	174-176	ZSM-A20111355	KF603955/CFAP029-11	
	SUB_4	ZSMA20111357_HT16	-51.086	-61.733	174-176	ZSM-A20111357	KF603956/CFAP030-11	
	SUB_4	ZSMA20111359_HT18	-51.086	-61.733	174-176	ZSM-A20111359	KF603957/CFAP031-11	
	SUB_4	ZSMA20111360_HT15	-51.086	-61.733	174-176	ZSM-A20111360	KF603958/CFAP032-11	
	SUB_4	ZSMA20111361_HT19	-51.086	-61.733	174-176	ZSM-A20111361	KF603959/CFAP033-11	
	SUB_5	HF26_027	-52.600	-73.640	19	ZSM-A20160452	KY272344	
	SUB_5	HF26_030	-52.600	-73.640	15-20	ZSM-A20160448	KY272343	
	SUB_5	HF26_059	-53.007	-73.923	31	ZSM-A20160457	KY272349	
	SUB_5	HF26_086	-53.357	-73.087	9	ZSM-A20160465	KY272341	KY272448
	SUB_5	HF26_120	-53.702	-72.041	22	ZSM-A20160472	KY272355	
	SUB_5	HF26_367	-53.357	-73.087	20	ZSM-A20160468	KY272351	KY272447
	SUB_5	HF26_368	-53.357	-73.087	18	ZSM-A20160467	KY272346	KY272449
	SUB_5	HF26_369	-53.357	-73.087	14	ZSM-A20160466	KY272353	
	SUB_5	HF26_373	-53.379	-73.159	14	ZSM-A20160488	KY272347	KY272453
	SUB_5	HF26_392	-53.379	-73.159	17	ZSM-A20160493	KY272345	KY272450
	SUB_5	HF26_439	-53.379	-73.159	17	ZSM-A20160483	KY272354	KY272452
	SUB_5	HF26_451	-53.379	-173.159	21	ZSM-A20160490	KY272352	KY272451
	SUB_5	HF26_458	-53.379	-73.159	14	ZSM-A20160494	KY272350	KY272455
	SUB_5	HF26_647	-53.896	-71.311	23	ZSM-A20160476	KY272342	
	SUB_5	HF26_648	-53.896	-71.311	23	ZSM-A20160477	KY272348	
	SUB_5	KT982315	-53.270	-66.386	96		KT982315	KY272456
	SUB_5	ZSMA20111340_HT12	-55.000	-68.315	24	ZSM-A20111340	KF603948/CFAP018-11	
<i>Pallenopsis yepayekae</i>	Pye.1	HF16_171	-50.338	-75.381	20	ZSM-A20119982	KY272271	
<i>P. yepayekae</i>	Pye.1	HF16_187	-50.359	-75.339	15	ZSM-A20119968	KY272266	KY272369
<i>P. yepayekae</i>	Pye.1	HF16_309	-50.359	-75.339	20	ZSM-A20119978	KY272252	
<i>P. yepayekae</i>	Pye.1	HF16_476_1	-50.353	-75.283	20	ZSM-A20119979	KY272287	
<i>P. yepayekae</i>	Pye.1	HF16_476_2	-50.353	-75.283	20	ZSM-A20160580	KY272283	
<i>P. yepayekae</i>	Pye.1	HF16_476_3	-50.353	-75.283	20	ZSM-A20160701	KY272256	KY272385
<i>P. yepayekae</i>	Pye.1	HF16_519_1	-50.412	-75.345	20	ZSM-A20160702	KY272262	KY272370
<i>P. yepayekae</i>	Pye.1	HF16_519_2	-50.412	-75.345	31	ZSM-A20160703	KY272265	KY272371
<i>P. yepayekae</i>	Pye.1	HF16_520	-50.412	-75.345	20	ZSM-A20119986	KY272260	
<i>P. yepayekae</i>	Pye.1	HF16_521_1	-50.412	-75.345	19	ZSM-A20119985	KY272261	
<i>P. yepayekae</i>	Pye.1	HF16_563	-50.359	-75.339	20	ZSM-A20160704	KY272258	
<i>P. yepayekae</i>	Pye.1	HF21_212	-45.661	-73.218		ZSM-A20160705	KY272286	KY272374
<i>P. yepayekae</i>	Pye.1	HF21_225	-45.921	-73.964		ZSM-A20160579	KY272270	
<i>P. yepayekae</i>	Pye.1	HF21_387	-45.763	-73.492	15	ZSM-A20160581	KY272251	
<i>P. yepayekae</i>	Pye.1	HF21_387_2	-45.763	-73.492	15	ZSM-A20160583	KY272285	KY272375
<i>P. yepayekae</i>	Pye.1	HF21_426_1	-45.521	-73.554	19	ZSM-A20160584	KY272255	
<i>P. yepayekae</i>	Pye.1	HF21_426_3	-45.521	-73.554	19	ZSM-A20160706	KY272253	KY272388
<i>P. yepayekae</i>	Pye.1	HF21_79	-45.662	-73.849	20	ZSM-A20160707	KY272257	KY272373
<i>P. yepayekae</i>	Pye.1	HF24_213	-53.007	-73.923		ZSM-A20160529	KY272268	KY272372
<i>P. yepayekae</i>	Pye.1	HF26_029	-52.600	-73.640	17	ZSM-A20160450	KY272281	
<i>P. yepayekae</i>	Pye.1	HF26_031	-52.600	-73.640	15-20	ZSM-A20160454	KY272274	

(Continued)

TABLE 1 | Continued

Species	Clade	Name	Lat	Lon	Depth	ZSM-Voucher Number	COI-GenBank/ BOLD Number	ITS-GenBank/ BOLD Number
<i>P. yepayekae</i>	Pye.1	HF26_032	-52.600	-73.640	15–20	ZSM-A20160447	KY272279	KY272376
<i>P. yepayekae</i>	Pye.1	HF26_090	-53.379	-73.159	14	ZSM-A20160499	KY272254	KY272384
<i>P. yepayekae</i>	Pye.1	HF26_264	-52.879	-74.350	20	ZSM-A20160460	KY272275	KY272386
<i>P. yepayekae</i>	Pye.1	HF26_265	-52.879	-74.350	20	ZSM-A20160459	KY272278	KY272377
<i>P. yepayekae</i>	Pye.1	HF26_306	-52.879	-74.350	25	ZSM-A20160458	KY272282	
<i>P. yepayekae</i>	Pye.1	HF26_363	-53.007	-73.923	20	ZSM-A20160462	KY272284	
<i>P. yepayekae</i>	Pye.1	HF26_376	-53.379	-73.159	16	ZSM-A20160527	KY272276	
<i>P. yepayekae</i>	Pye.1	HF26_378	-53.379	-73.159	29	ZSM-A20160498	KY272277	KY272387
<i>P. yepayekae</i>	Pye.1	HF26_394	-53.379	-73.159	17	ZSM-A20160484	KY272264	KY272389
<i>P. yepayekae</i>	Pye.1	HF26_396	-53.379	-73.159	17	ZSM-A20160497	KY272259	KY272381
<i>P. yepayekae</i>	Pye.1	HF26_397	-53.379	-73.159	17	ZSM-A20160496	KY272272	KY272382
<i>P. yepayekae</i>	Pye.1	HF26_555	-53.702	-72.041	21	ZSM-A20160473	KY272263	KY272380
<i>P. yepayekae</i>	Pye.1	HF26_562	-53.896	-71.311	23	ZSM-A20160478	KY272267	KY272378
<i>P. yepayekae</i>	Pye.1	HF26_563	-53.818	-71.056	23	ZSM-A20160479	KY272280	KY272379
<i>P. yepayekae</i>	Pye.1	HF26_566	-53.818	-71.056	7	ZSM-A20160481	KY272269	KY272383
<i>P. yepayekae</i>	Pye.1	HF26_601	-53.587	-72.338	16	ZSM-A20160471	KY272273	
<i>P. yepayekae</i>	Pye.1	ZSMA20111000_HT07	-48.737	-75.415	15	ZSM-A20111000	KF603944/CFAP013-11	
<i>P. yepayekae</i>	Pye.1	ZSMA20111002_HT06	-50.835	-74.139	25	ZSM-A20111002	KF603947/CFAP017-11	
<i>P. yepayekae</i>	Pye.1	ZSMA20111005_HT04	-48.737	-75.415	23	ZSM-A20111005	KF603945/CFAP014-11	
<i>P. yepayekae</i>	Pye.1	ZSMA20111006_HT01	-43.418	-74.081	20	ZSM-A20111006	KF603941/CFAP007-11	
<i>P. yepayekae</i>	Pye.1	ZSMA20111016_HT09	-48.608	-74.899	32	ZSM-A20111016	KF603943/CFAP012-11	
<i>P. yepayekae</i>	Pye.1	ZSMA20111339_HT05	-43.775	-73.029	19	ZSM-A20111339	KF603949/CFAP019-11	
<i>P. yepayekae</i>	Pye.2	ZSMA20111003_HT03	-43.418	-74.081	25	ZSM-A20111003	KF603940/CFAP006-11	
<i>P. yepayekae</i>	Pye.2	ZSMA20111004_HT01	-43.410	-74.084	9	ZSM-A20111004	KF603939/CFAP005-11	
<i>P. yepayekae</i>	Pye.2	ZSMA20111009_HT02	-43.393	-74.132	26	ZSM-A20111009	KF603938/CFAP004-11	
<i>P. yepayekae</i>	Pye.2	ZSMA20111012_HT08	-43.771	-73.044	22	ZSM-A20111012	KF603942/CFAP008-11	
<i>P. pilosa</i>	OG	PxxE_001	-54.350	3.193		ZSM-A20160732	KC794967	KY272459
<i>P. pilosa</i>	OG	PxxE_002	-54.213	-32.606	200	ZSM-A20160733	KC794968	KY272460
<i>P. pilosa</i>	OG	PxxE_003	-54.397	3.521		ZSM-A20160734	KY272362	
<i>P. pilosa</i>	OG	PxxE_005	-54.397	3.521		ZSM-A20160735	KY272361	
<i>P. macronyx</i>	OG	PS42_164_2	-62.133	-57.667		ZSM-A20160619	KY272364	
<i>P. macronyx</i>	OG	PS42_164_3	-62.133	-57.667		ZSM-A20160620	KY272363	
<i>P. macronyx</i>	OG	PS42_164_4	-62.133	-57.667		ZSM-A20160621	KY272365	

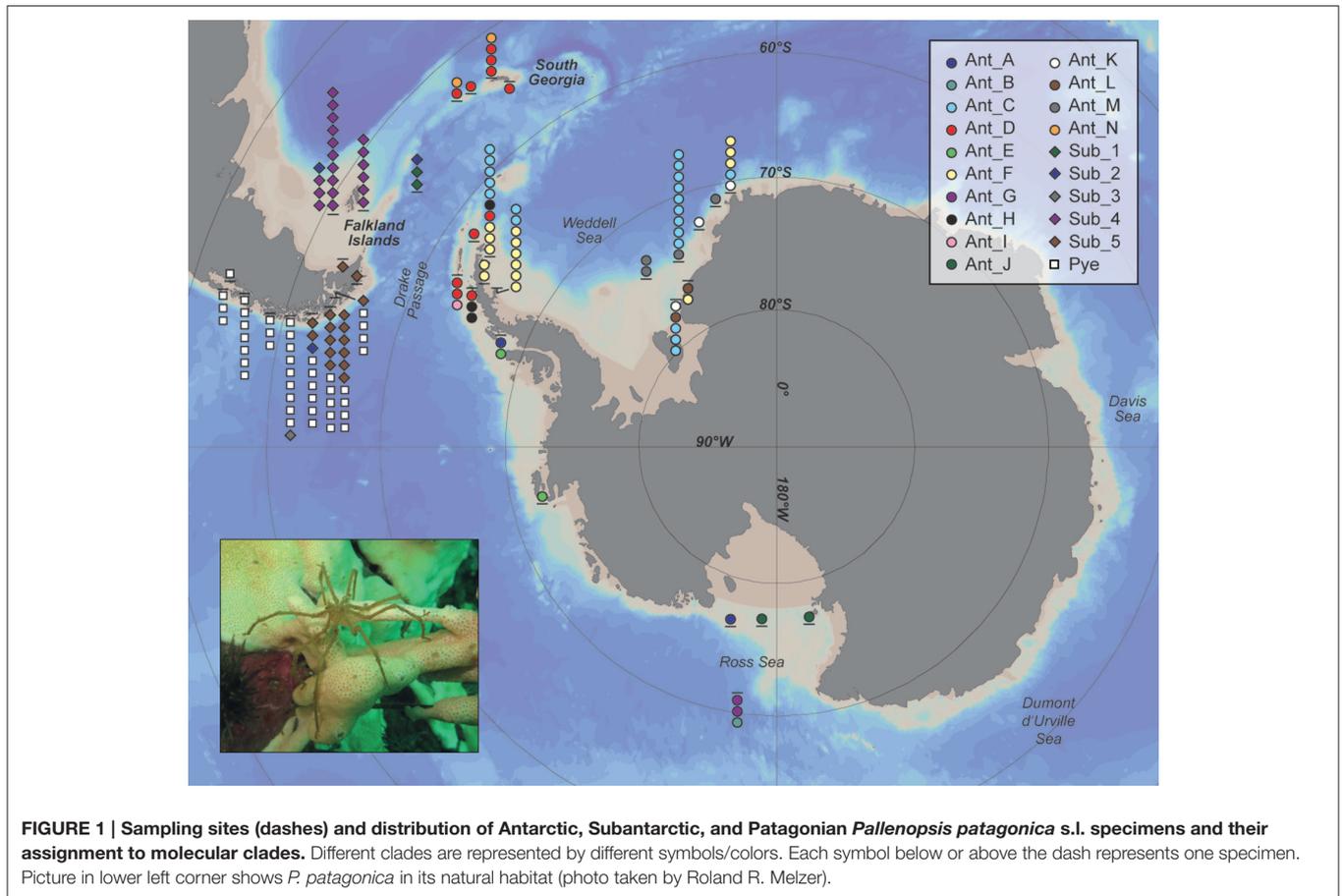
Center for Polar and Marine Research, Bremerhaven, Germany). After collection, specimens were stored in ethanol (96%). Specimens were morphologically inspected and assigned to *P. patagonica* s.l. before being molecularly studied.

Molecular Analyses

Muscle tissue was extracted from the tibia using sterile scalpel and forceps. DNA was isolated from the tissue using a modified salt precipitation protocol after Sunnucks and Hales (1996; see Weiss and Leese, 2016). Extracted DNA was eluted in 100 μ l TE minimum buffer (1 mM Tris BASE, 0.1 mM EDTA, pH 8.0). The amplification of the mitochondrial cytochrome c oxidase subunit I gene (COI) and a ribosomal gene region covering the 18S–ITS1–5.8S–ITS2–28S stretch (ITS) was carried out in 25 μ l reactions containing 1x (2.5 μ l) PCR buffer (5Prime),

0.2 mM dNTPs, 0.5 μ M of each primer, 0.025 U/ μ l (0.125 μ l) Hotmaster Taq (5Prime) and 1 μ l template DNA, topped up to 25 μ l with sterile water. A 658 bp long fragment of the COI was amplified using the common barcoding primer pair HCO2198 and LCO1490 (Folmer et al., 1994). The optimal temperature profile for the PCRs with these primers was an initial denaturation at 94°C for 2 min, followed by 36 cycles of denaturation at 94°C for 20 s, annealing at 46°C for 30 s, extension at 65°C for 60 s, and a final extension at 65°C for 7 min.

For ITS, an approximately 1000 bp long fragment was amplified using primers ITSRA2 and ITS2.2 (Arango and Brenneis, 2013). PCR cycling program was initial denaturation at 94°C for 3 min, followed by 35 cycles of denaturation at 94°C for 30 s, annealing at 55°C for 75 s, extension at 65°C for 1 min, with a final extension at 65°C for 5 min.



For sequencing, 10 U (0.5 μ l) Exonuclease I (Thermo Scientific), 1.5 U (1 μ l) FastAP Thermosensitive Alkaline Phosphatase (Thermo Scientific) and 9 μ l PCR product per reaction were used. The purification mix was incubated for 25 min at 37°C, followed by a denaturation step at 85°C for 15 min. For sequencing at GATC Biotech AG (Cologne, Germany) 5 μ l of purified PCR product was mixed with 5 μ l of 5 pmol/ μ l primer. Forward and reverse primers were used to sequence both directions of the DNA strands.

For ITS sequences of samples reported by Harder et al. (2016; herein labeled with GenBank numbers starting with KT98) DNA extraction was performed as stated in Harder et al. (2016). For ITS amplification the same primer pair as mentioned above was used. PCR mixture consisted of 1x PCR buffer, 0.75 U Taq DNA polymerase (5Prime, Hotmaster Taq), 2.5 mM Mg²⁺, 10 nmol of each dNTP, 1 μ l of template DNA, 0.5 μ M of each primer, and water to 25 μ l. PCR cycling program was run, with an initial denaturation at 94°C for 2 min, followed by 37 cycles of denaturation at 94°C for 20 s, annealing at 55°C for 30 s, extension at 65°C for 80 s, with a final extension at 65°C for 10 min. Successful amplification was confirmed by visualizing PCR products on a 1% agarose gel stained with ethidium bromide. Target PCR product was gel extracted and purified using a Qiagen QIAquick[®] Gel Extraction Kit according to the manufacturer's recommendations. Bidirectional Sanger

sequencing of amplicons was performed at High Throughput Genomics Center (Seattle, WA, USA).

Phylogenetic Analyses

For COI, *P. patagonica* s.l. sequences from Weis et al. (2014; $n = 34$ including five downloaded from NCBI) and Harder et al. (2016; $n = 26$) were added to the final data set. ITS sequences of specimens of both previous studies were generated and also included in the ITS alignment, too.

For both gene regions, sequences were edited with Geneious v. 8.1.3 (Kearse et al., 2012) and aligned in Geneious using MAFFT v. 7.017 Multiple Sequence Alignment (Katoh and Standley, 2013) with default parameters as implemented in Geneious, with a gap opening penalty of 1.53 and offset value of 0.123. For COI, sequences were translated into amino acids using the invertebrate mitochondrial genetic code (transl_table=5) to verify that all codons could be translated without stop codons. For ITS, a version of the alignment where ambiguously aligned regions were removed was produced with Gblocks v. 0.91b (Castresana, 2000) using less stringent parameters (smaller blocks, gaps in final alignment allowed, less strict flanking positions) as has been done in Dietz et al. (2015b). For analyses when only unique copies were needed, sequences were collapsed into unique sequences ("haplotypes" for COI data) with the online tool FaBox v. 1.41 (Villesen, 2007).

For both data sets a maximum-likelihood (ML) analysis was performed with RAxML v. 8.2.4 (Stamatakis, 2014) using the GTRCAT model of sequence evolution and branch support was assessed with 10,000 rapid bootstrap replicates. In addition, for the mitochondrial data set a resolved ultrametric gene tree was calculated using BEAST v. 1.8.3 (Drummond et al., 2012) with the model specified by jModelTest v. 2.1.10 (Guindon and Gascuel, 2003; Darriba et al., 2012). An XML file was created with BEAUti v. 1.8.3 (Drummond et al., 2012) with the following settings: HKY+G+I as substitution models and 80×10^6 as length of MCMC chain sampling every 1000th tree. Convergence of the likelihood and appropriate effective sampling size (ESS > 200) of parameter estimates were checked using TRACER v. 1.6 (Rambaut et al., 2014), and a consensus tree was calculated using TREEANNOTATOR v. 1.8.3 of the BEAST package. Furthermore, uncorrected pairwise distance matrices were created using MEGA v. 7 (Tamura et al., 2011).

Species Delimitation Methods

For species delimitation analysis of the COI data set we used ABGD (Automatic Barcode Gap Discovery; Puillandre et al., 2012). As no clear barcode gap was found in the pairwise distance data, ABGD results varied strongly depending on single sequences and run parameters tested. Results presented here are mainly from the default settings but using Kimura-2-parameter (K2P) distance correction. The same settings were applied to the ITS alignment (including and excluding ambiguously aligned regions). Due to the smaller data set for the ITS alignment and the fact that informative alignment gaps cannot easily be interpreted as additional character in tree-based delimitation methods, further species delimitation methods were only applied to the COI data set. The final mitochondrial COI ML tree was used to perform a Bayesian Poisson Tree Processes (bPTP) analysis using the web server (<http://species.h-its.org/ptp>; Zhang et al., 2013). Furthermore, a Generalized Mixed Yule Coalescent (GMYC) analysis based on the resolved ultrametric gene tree was conducted at the web server (<http://species.h-its.org/gmyc>; Fujisawa and Barraclough, 2013) using the single-threshold method only (see Fujisawa and Barraclough, 2013).

Molecular Clock Analysis

A calibrated molecular clock rate for sea spiders has not been reported in previous studies. However, in order to infer possible divergence date ranges for the different clades we applied a widely adopted COI molecular clock rate reported for insects: 1.15% per myr and lineage (Brower, 1994). BEAST v. 1.8.2 was used to estimate divergence times using an HKY+I+G evolution model as well as an uncorrelated local clock model. Analyses were run for 10×10^6 generations sampling every 1000th tree. Convergence of parameter estimates and ESS control and subsequent steps were done as described above.

RESULTS

Number of COI Clades

The data set of *P. patagonica* s.l. was extended to a total of 173 specimens including 47 sequences from *P. yepayekae*. For the first

time, we studied individuals from the Strait of Magellan and the Eastern Weddell Sea. Further sequences from *P. pilosa* (Hoek, 1881) ($n = 3$) and *P. macronyx* (Bouvier, 1911) ($n = 3$) were added to the data set as outgroups.

The final COI alignment consisted of 426 bp (GC content 32.9%) with 278 identical and 128 parsimony informative sites. Neither stop codons nor frame shift mutations were observed after translation. Both the ML and Bayesian phylogenetic tree (Figure 2) resolved *P. patagonica* s.l. as monophyletic and well-separated from the outgroup. In addition, *P. yepayekae* represented a monophyletic group within *P. patagonica* s.l. Moreover, all individuals from the Antarctic shelf formed an “Antarctic super-clade” that also contained one clade with specimens from South Georgia, i.e., one of the Subantarctic islands. The two other specimens collected around South Georgia that did not cluster inside this group represented the basal-most group (Clade N) in the whole *P. patagonica* s.l. group (Figure 2). Analysis of the final COI alignment with ABGD using K2P substitution model revealed a steady decrease from 21 to 13 in number of recovered groups between $P = 0.002$ and $P = 0.05$ in the recursive partition. No clear barcode gap was visible when plotting pairwise uncorrected distances between *P. patagonica* s.l. specimens (Supporting information Table S1; see upper diagram in Figure 3 showing distances between members of the Antarctic clade). When choosing a threshold value of $P = 0.05$ ABGD suggested 13 clades, with several formerly reported clades merged (clade E, F, and G from Harder et al., 2016). At $P = 0.06$ ABGD merged all sequences into one group. Analysis of the Bayesian tree with bPTP, suggested the presence of 20 distinct groups, hence, subdivided five ABGD groupings further resulting in seven additional clades. With 22 groups, GMYC reported the highest number of clades for the ML tree. In contrast to bPTP, GMYC furthermore subdivided *P. yepayekae* and clade ANT_D into two subclades each. Here, we named the clades according to the bPTP results (see Section Discussion for further information). Several “clades” (two for ABGD, four for bPTP and GMYC) consisted of single specimens only (Figure 2).

Upon reviewing the data set, the following points are of particular interest to address our hypotheses. Newly collected specimens from the Strait of Magellan that were morphologically determined as *P. yepayekae* clustered with available sequences of *P. yepayekae* (Weis et al., 2014; Figure 2). Using the GMYC delimitation method, this species was split into two subclades. One subclade (Pye.2, Table 1) included all three haplotypes reported for four specimens sampled in the Chilean region Los Lagos, i.e., at the northernmost occurrence of *P. yepayekae*. Both other delimitation methods resolved *P. yepayekae* as a single clade.

All *P. patagonica* samples from the Strait of Magellan clustered together with sequences that in Weis et al. (2014) formed a sister clade to the Falkland clade (specimens ZSMA20111017 and ZSMA20111340, see Figure 2 in Weis et al., 2014). Average p-distance between this clade and the Falkland clade is 2.7%. The 15 new samples collected around South Georgia clustered together with a specimen from Shag Rocks that was reported as member of the Antarctic clade of *P. patagonica* (PpaE_001, see Weis et al., 2014). All these specimens formed a subclade (ANT_D.2,

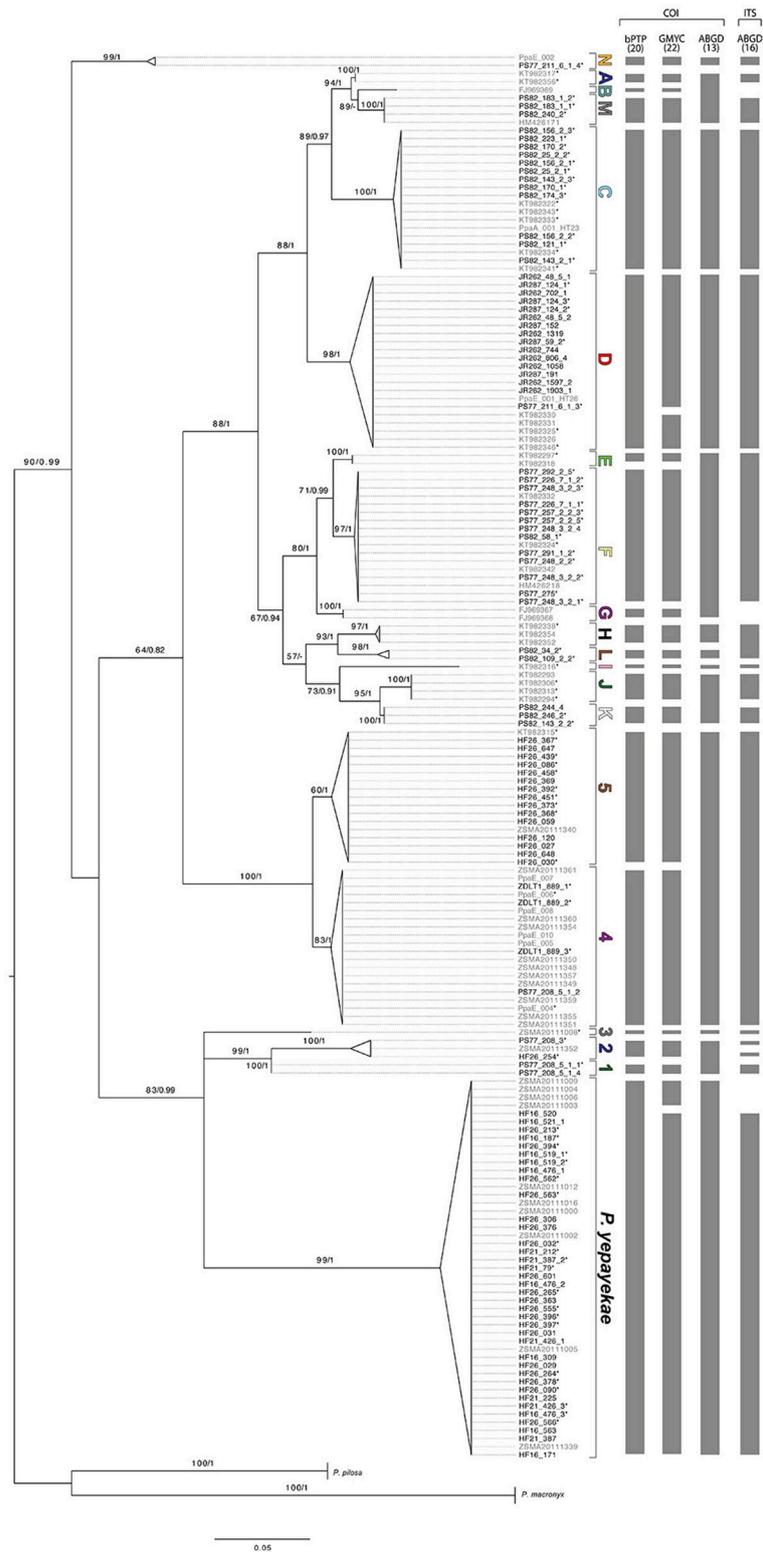


FIGURE 2 | Maximum likelihood tree of COI sequences of *Pallenopsis patagonica* s.l. combining new (black) and previously (gray) reported samples. *P. pilosa* and *P. macronyx* serve as outgroups. Asterisks (*) indicate availability of ITS sequence data for the respective specimens. Bootstrap/posterior probabilities values above 50/0.5 are provided next to each node. Letters and numbers stand for mitochondrial clades from Antarctica (ANT) and the Subantarctic (SUB), respectively. Columns show results of COI-based species delimitation methods (bPTP, GMYC, and ABGD), number in parentheses denote the total number of predicted species by each method. In addition, results of ABGD when analyzing ITS (see **Figure 4**) are shown in the rightmost column.

Table 1) of clade ANT_D that so far consisted of specimens sampled around the tip of the Antarctic Peninsula (ANT_D.2; Harder et al., 2016). One sample from Shag Rocks grouped together with one individual from South Georgia (PpaE_002), which form the most basal clade (ANT_M) within *P. patagonica* s.l. For *P. patagonica* s.l. from the Antarctic shelf, our data set, analyzed with bPTP and GMYC, revealed the presence of three further clades (ANT_K, ANT_L, and ANT_M) in addition to the 10 clades reported from the Antarctic by Harder et al. (2016). These three new clades are exclusively found in the Eastern Weddell Sea (**Figure 1**). The newly found clade ANT_K is sister clade to ANT_J, clade ANT_L to ANT_H, and clade ANT_M to ANT_B (**Figure 2**). The two sequences HM426171 and HM426218 reported in Weis et al. (2014) as *P. buphtalmus* (Pushkin, 1993) and *P. latefrontalis* (Pushkin, 1993) based on provisional ID in BOLD by that time, clustered together with clade ANT_M and clade ANT_F, respectively. Sampling sites of these are also in the same region.

Nuclear Support for COI Based Species Delimitation

We tested for congruence between the COI and the nuclear ITS clades by analyzing 96 sequences from the majority (bPTP: 18 of 20; GMYC 19 of 22) of reported COI clades (**Figure 2**). Very few ambiguities were included in sequences due to sequence quality issues for some sequences. The ITS sequence for JR287_124_2 was composed only of two shorter single read sequences (forward and reverse) but had no overlap (108 missing data symbols, “?”). The initial alignment was 1071 bp long, but was shortened to 583 bp after filtering for noisy positions with GBLOCKS. The final alignment consisted of sequences of 344–582 bp in length with 126 sites being identical and 159 parsimony informative. The base composition was very homogenous with A: 23.2%, C: 27.2%, G: 25.8%, and T: 23.8%. The number of ITS haplotypes was 23 representing 18 of the mitochondrial bPTP clades, i.e., four mitochondrial clades (ANT_C, ANT_F, SUB_2, and SUB_5) had two corresponding ITS sequences and *P. yepayekae* had three. ANT_H and ANT_L shared the same sequence. No heterozygous individuals were observed.

The phylogenetic ITS tree is much less resolved than the COI tree (**Figure 4**). However, most samples grouped similar to the COI tree. Separated, albeit poorly supported in the ITS tree, ANT_E and ANT_F clustered together when analyzed with ABGD. Pairwise identity between sequences of the clades was very high (98.9%). Together with ANT_G, for which no ITS data could be obtained, these clades represented a monophyletic group in the mitochondrial tree. In few cases there were minor disagreements in terms of the resolved clades. For examples ANT_L and ANT_H have identical sequences for ITS and hence grouped together. In the COI tree, these two clades represented slightly divergent sister clades. The other dissimilarity between the two phylogenetic trees was a well-supported nuclear clade (bootstrap support of 99%) that included ANT_A and ANT_D as sister groups (no shared haplotypes, however). ABGD distinguished the groups but in

the mitochondrial tree ANT_A and ANT_D were not sister groups, yet closely related. In general, ITS showed substantially less variation than COI, and most mitochondrial clades were supported by ITS with minor exceptions mentioned above. However, within the clade SUB_2 we obtained two different ITS sequences from two individuals (PS77_208_3 and HF26_254) that showed much greater nucleotide variation within the ITS than the COI data. Interestingly, in the ITS tree they formed a paraphylum rather than a monophylum as the sequence of another clade (SUB_1) was also included. ABGD split these sequences with low pairwise identity (90.5%) into two separate groups.

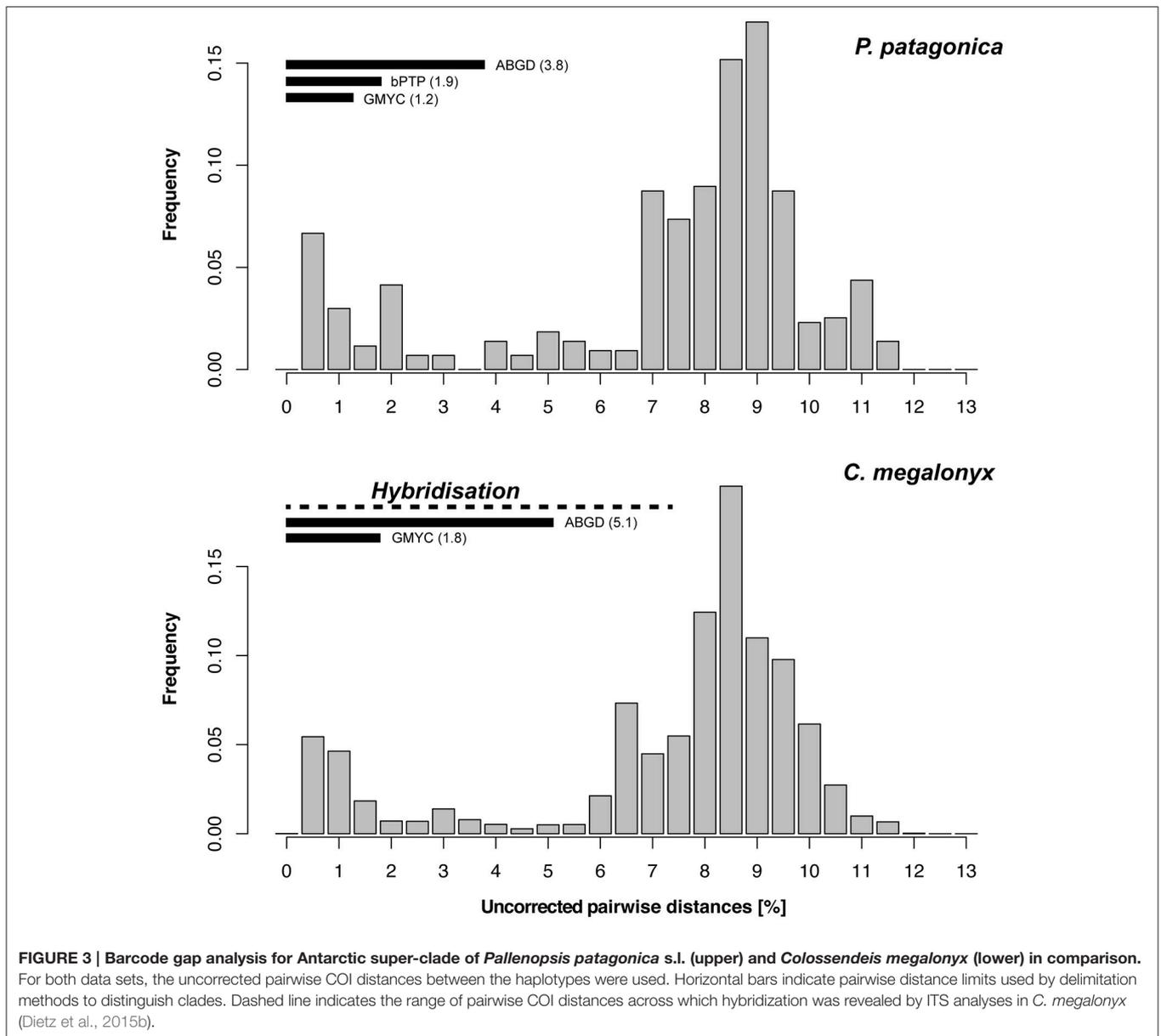
Biogeographic Patterns

The new samples included here prove that *P. yepayekae* also occurs in the Strait of Magellan and here even in sympatry with another clade of *P. patagonica* s.l. (SUB_5; see **Figure 1**). SUB_5 was represented by only two specimens in Weis et al. (2014) and in contrast to Weis et al. (2014) merged into the “Falkland clade” in Harder et al. (2016). Here, with the additional data, both bPTP and GMYC supported that SUB_5 represents a distinct clade. One formerly reported *P. patagonica* s.l. clade (termed HT25 in Weis et al., 2014), represented by a single specimen found around South Georgia (Ppa_E002), now included one further specimen from the Shag Rocks (PS77_211_6_1_4). This clade is herein referred to as ANT_N. Vice versa, another clade of *P. patagonica* s.l., formerly represented by a single specimen from Shag Rocks (Ppa_E001), now clustered together with newly collected specimens from South Georgia (representing one subclade of ANT_D). ANT_N formed a cluster basal to the split between the Antarctic super-clade (i.e., specimens sampled South of the Polar Front) and the Falkland/Strait of Magellan clade (**Figure 2**). South Georgia individuals belonging to ANT_D thus grouped within the Antarctic clade reported by Weis et al. (2014) and clade D reported by Harder et al. (2016). Hence, these two clades (ANT_N and ANT_D) that occur in the same area, are not sister clades but only distantly related.

The new specimens sampled from the Eastern Weddell Sea grouped into five clades. Two of these clades, ANT_C and ANT_F, were already reported from the Antarctic Peninsula by Harder et al. (2016). Thus, our new data extended the reported distribution range for these two clades to the Eastern Weddell Sea. Specimens of the other three clades have not been reported earlier and were only found in the Eastern Weddell Sea. Specimens of clade ANT_D, found at the northernmost tip of the Antarctic Peninsula, grouped together with the individuals mentioned above from around South Georgia and Shag Rocks (**Figure 2**). No clades with individuals from either side of the Antarctic Polar Front were found in our data set.

Divergence Dating

The divergence from the most recent common ancestor of *P. patagonica* s.l. occurred 13.6 myr before present [HPD 95% interval: 9.8–17.7 myr before present (BP)]. Also, the divergence of the Antarctic vs. the Falkland/Magellan clade took place in the mid Miocene (9.5 myr BP), 7.2–12.9 myr BP). Divergence of the distinct mitochondrial clades occurred (independently



on whether choosing ABGD, bPTP or GMYC as a delimiting criterion) in the Plio- and Pleistocene, mostly before the last 2 myr BP (Supplementary Figure S1).

DISCUSSION

Number of Mitochondrial Clades

As predicted by the first hypothesis, we found additional mitochondrial clades within *P. patagonica* s.l. when analyzing the extended COI data set with bPTP and GMYC. The groupings of bPTP and GMYC were congruent for most clades with the exception that GMYC further subdivided *P. yepayekae* and ANT_D into two geographically separated subclades each. However, when using ABGD with default settings, the

number of mitochondrial clades inferred was actually smaller than the number reported by Harder et al. (2016) (Figure 2, Supplementary Table S2).

Which Mitochondrial Clades Can Be Considered As Species?

When trying to find an objective value that best describes the number of species (defined as independently evolving units) with the classical COI barcoding alone, the original approach was to quantify intra- vs. inter-specific genetic distances through a barcoding gap analysis that defines the maximum threshold distance found within a species. For animal taxa, this value has often been found at 2% pairwise distances (e.g., Hebert et al., 2003). Other approaches expect distinct species

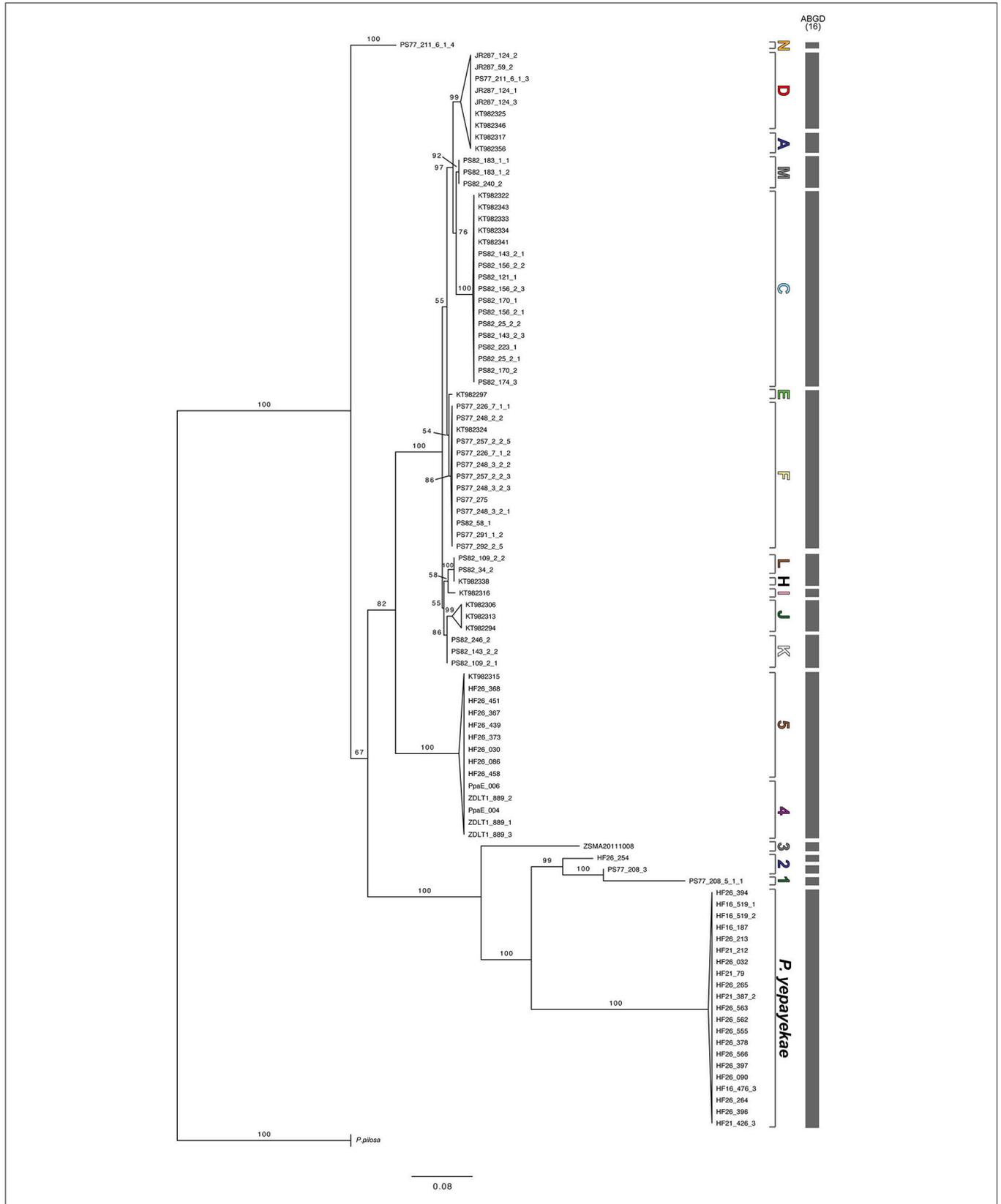


FIGURE 4 | Maximum likelihood tree of ITS sequences of *Pallenopsis patagonica* s.l. Bootstrap values above 50 are provided next to each node. Letters on the right correspond to the labels used in the mitochondrial tree (see **Figure 2**). Letters and numbers stand for mitochondrial clades from Antarctica (ANT) and the Subantarctic (SUB), respectively. Bars represent results of the ABGD analysis that was based on the full alignment.

to show a 10x greater divergence than found within species (Hebert et al., 2004). Barcode gap analyses within sea spiders have reported intraspecific threshold distances of up to 5% (Mahon et al., 2008). Here, we could not detect a distinct barcoding gap for the *P. patagonica* species complex but rather a gradient of pairwise distances, though many at low frequency (Supporting Information Table S1). This is similar to results observed within the sea spider complex *C. megalonyx* (Dietz et al., 2015b). Consequently, species delimitation based on COI is not straightforward and we followed several lines of argumentation summarized in Kekkonen and Hebert (2014) in order to discuss whether mitochondrial clades resemble species or not. According to the ideas presented in Kekkonen and Hebert (2014), species status can be assigned to clades with a full match of all different species delimitation methods. This is the case for the following six clades: SUB_3, ANT_C, ANT_H, ANT_I, ANT_L, and ANT_N. For all other clades, only a partial match between the different methods was observed. In such cases, Kekkonen and Hebert (2014) suggest to test whether (a) the resolved clades are monophyletic, (b) individuals are supported by diagnostic characters (nucleotide substitutions, insertions or deletions, see also Jörger and Schrödl, 2013), or (c) specimens of different clades occur in sympatry. All three criteria are based on species concepts. Both, monophyletic entities and diagnostic characters missing in sister taxa matter for the phylogenetic species concept. The biological species concept requires groups that are reproductively isolated, which in nature can only be detected when groups occur in sympatry. Due to a limited sample size and geographical range and hence the potential for unsampled haplotypes leading to ascertainment biases, the criterion of diagnostic characters is not considered here and only the two remaining criteria, monophyly and occurrence in sympatry, are applied to the evaluation of the mitochondrial data set.

The GMYC-subclades for *P. yepayekae* are the only example of the data set where the monophylum criterium cannot be applied, because although the individuals from Los Lagos themselves form a monophylum, the remaining specimens, which represent the majority, would be rendered as paraphyletic. Furthermore, specimens of both subclades do not occur in sympatry. As the Los Lagos specimens represent the northernmost occurrence of *P. yepayekae* known to date, a straightforward explanation of this pattern is isolation-by-distance. In particular because genetic differences between specimens from these two clades (max. uncorrected p-distances observed 0.9%) lie well within the range typically reported as intraspecific for other sea spider species (Mahon et al., 2008; Krabbe et al., 2010; Arango and Brenneis, 2013; Dietz et al., 2015a,b) as well as other arthropods (see Supporting Information Table S1 in Smith et al., 2005) we refrain from assigning species-level status to these two subclades and rather accept the grouping based on bPTP and ABGD.

For clade ANT_D, the found divergence of 1.6% between the two GMYC-subclades (ANT_D.1 vs. ANT_D.2, Antarctic Peninsula vs. South Georgia, respectively) is larger than between the GMYC-subclades of *P. yepayekae*. But because this value is still within the range reported as intraspecific and the subclades occur in different regions it cannot be ruled out that they

represent two geographically separated populations. Hence, we refer to these subclades as one clade. ANT_D represents the only reported *P. patagonica* s.l. clade that crossed the deep sea between the continental shelf and the Subantarctic islands, but stayed within the Polar Front. Gene flow between the Antarctic continental shelf and South Georgia has already been reported in a few studies on other benthic invertebrates (Thornhill et al., 2008; Wilson et al., 2009; Dietz et al., 2015a,b).

ABGD merged five of the bPTP/GMYC-clades. All single bPTP/GMYC-clades are reciprocally monophyletic, but only SUB_1 and SUB_2 occur in sympatry. Following the protocol of Kekkonen and Hebert (2014), SUB_1 and SUB_2 would represent two distinct species as revealed by bPTP and GMYC. All other clades merged by ABGD do not occur in sympatry and therefore species assignments are not possible based on the limited data set. ANT_A, ANT_B, and ANT_M were sampled from Ross Sea and Eastern Weddell Sea with not exceedingly high uncorrected pairwise distances ranging from 2.1 to 3.5%. Thus, the grouping suggested by ABGD seems adequate. ANT_E, ANT_F, and ANT_G were sampled from the Western side of the Antarctic Peninsula, on both sides of the Weddell Sea and the Ross Sea. This pattern could be the result of isolation by distance, too. Harder et al. (2016) also found clade G as a distinct clade using bPTP, GMYC, and ABGD as delimitation methods. Clades E and F were separated by bPTP and GMYC, but merged with ABGD. Although Harder et al. (2016) decided to keep all three distinct, we suggest to be more careful here in particular in view of the few specimens available (only two for each of clade E and G) and the shallow divergences. Similar as above, ANT_J and ANT_K from the Ross Sea and Weddell Sea, respectively, have a moderate uncorrected pairwise distance of 2.1%. Hence, they are also not treated as different units here. This is also true for groupings of SUB_4 and SUB_5. Specimens of both again do not occur in sympatry (Strait of Magellan vs. Falkland Plateau), and divergence falls well in the range of values reported as intraspecific (average uncorrected pairwise distance 1.93%). In the reported cases of (partial) mismatch between the three delimitation methods it is difficult to apply a general rule, because no clear barcoding gap is known that allows for a clear cut between intra- and inter-specific genetic distances. The fact that we find clades, e.g., ANT_A and ANT_C, with very low intra-clade divergences (<1%) despite a broad distribution range would suppose that intraspecific genetic distances are small also for species with a broad distribution range (i.e., argument against isolation-by-distance). However, limited sampling size does not allow for further conclusions.

We suggest that for partial matches between delimitation methods every case should be evaluated on its own. Combining all these arguments in a conservative way, we suggest 14 distinct evolutionary units in *P. patagonica* s.l. based on the COI data (Table 2, Supporting information Table S2).

Nuclear Support for COI Based Species Delimitation

As shown in Dietz et al. (2015b) for pycnogonids only looking at mitochondrial data can lead to overestimation and

TABLE 2 | Comparison of results from species delimitation analyses using mitochondrial (COI) and nuclear (ITS) data of *Pallenopsis patagonica* and a final recommendation for groupings.

Clade	COI (14/22)	ITS (16/19)	Final grouping (15)
ANT_A			
ANT_B		NA	ANT_ABM
ANT_M			
ANT_C			ANT_C
ANT_D1			ANT D
ANT_D2			
ANT_E			ANT_EFG
ANT_F			
ANT_G		NA	
ANT_H			ANT HL
ANT_L			
ANT_I			ANT_I
ANT_J			ANT_J
ANT_K			ANT_K
ANT_N			ANT_N
SUB_1			SUB_1
SUB_2			SUB_2.1 SUB_2.2
SUB_3			SUB_3
SUB_4			SUB 4 + 5 (Falkland)
SUB_5			
Pye.1			<i>P. vepavekae</i>
Pye.2		NA	

Supporting Information Table S2 for a detailed list.

misinterpretation of the actual species number (see Toews and Brelford, 2012 for a review). Dietz et al. (2015b) also showed that ITS is a suitable marker for sea spiders as unlike the situation reported for other organisms (e.g., Weitemier et al., 2015) no multiple intragenomic variants for this gene could be detected when using high-throughput sequencing data (Leese et al., 2012). Comparing ITS and COI data we first see that contrary to *C. megalonyx* (Dietz et al., 2015b) no mito-nuclear discordances are observed in *P. patagonica* s.l. (Figures 2, 4) This indicates that different processes acted after initial mitochondrial lineage sorting on both sea spider species complexes and will be discussed below. Most importantly, ITS sequences are not shared between different COI clades in *P. patagonica*, with the exception of the mitochondrial sister clades ANT_H and ANT_L (both with a full match when comparing delimitation methods) that show the same ITS sequence. Even though ANT_E and ANT_F do not share one haplotype, ITS sequences are very similar and species delimitation analysis clusters them together. However, the groupings ANT_H and ANT_L as well as ANT_E and ANT_F (including ANT_G) represent a monophylum within the mitochondrial tree each. Whereas, ANT_H and ANT_L both had a full match when comparing mitochondrial data across delimitation methods, ANT_E and ANT_F were grouped together with ANT_G by

ABGD. Unfortunately, we were not able to obtain ITS sequences of an individual from ANT_G to analyse whether ANT_G also groups with ANT_E and ANT_F when analyzing ITS. However, ANT_E and ANT_F clustered into one ABGD group in the mitochondrial tree similar to the ITS tree. Given the lack of resolution, we here suggest not proposing species status for clades ANT_H, ANT_L, ANT_E, ANT_F and ANT_G based on mitochondrial results but suggest to refer to the two groups containing ANT_H and ANT_L as well as ANT_E, ANT_F and ANT_G as one clade each. We are aware of the fact that speciation could be recent and thus has not been picked up with ITS (Table 2).

Mitochondrial clades considered as one evolutionary unit sometimes comprise up to three different but genetically very similar ITS sequences. These sequences, however, cluster together when using ABGD on the ITS data set. For example, based on COI data clades ANT_J and ANT_K that were both distinguished as separate clades by bPTP and GMYC were treated as one hyper-clade as there was a lack of characters distinguishing them and ABGD clustered them together when analyzing mitochondrial data. But ITS data within this hyper-clade can be assigned to the two different bPTP/GMYC clades. The same holds true for further combinations that are also congruent between ITS and the bPTP/GMYC delimitation of the COI gene (SUB_1 and SUB_2, SUB_4, and SUB_5 as well as ANT_A and ANT_M). In the case of ANT_D, ITS sequences of all available representatives are identical. Thus, the assignment of all individuals to one clade by ABGD and bPTP is congruent with the ITS result. The more resolved delimitation into subclades suggested by the GMYC analysis of the COI data is not supported by the ITS data (Supporting Information Table S2). As above, we here also suggest a conservative approach to not treat these clades as distinct species prior to further evidence.

In view of the evidence from the COI data set and the protocol by Kekkonen and Hebert (2014) as well as the nuclear gene marker results we propose 15 putative evolutionary units for the current data set of *P. patagonica* s.l. (Table 2). The number is likely to change should further data become available as major regions of the Southern Ocean, especially East Antarctica, still remain unexplored. More important though, is to add further evidence that helps defining a clear boundary between intra- and interspecific characters of the species complex. COI combined for the first time with ITS data of *P. patagonica* s.l. is a major step forward. However, data are not sufficient for a final delimitation across all clades and additional independent characters (morphology, further genes) are needed to make clear statements. Still, the finding of mito-nuclear agreement supports that in contrast to *C. megalonyx* we can describe the distinct groups contained within *P. patagonica* s.l. reasonably well with the current data available.

Distribution Ranges

Our data supports a strong barrier effect of the Antarctic Polar Front as we did neither observe sister clades nor clades containing individuals from either side of the Polar Front of *P. patagonica* s.l. Such a pattern has also been observed for many

other benthic invertebrates (e.g., Page and Linse, 2002; Thornhill et al., 2008; Krabbe et al., 2010). The fact that individuals from South Georgia, i.e., a Subantarctic island south of the Polar Front, form one clade with individuals reported from the northernmost tip of the Antarctic Peninsula (ANT_D) and cluster within the Antarctic super-clade of *P. patagonica* s.l. hints at a colonization event of South Georgia from the Antarctic. This direction of gene flow makes sense as it is consistent with a pattern of colonization with the Antarctic Circumpolar Current from West to East (Leese et al., 2010). However, for *C. megalonyx* also shared haplotypes between South Georgia and the Antarctic Peninsula were found for one clade (Clade A; Dietz et al., 2015b). Interestingly, here genetic diversity patterns clearly indicated the opposite pattern, i.e., gene flow from South Georgia to the tip of the Antarctic Peninsula. Dietz et al. (2015b) considered South Georgia as the likely refugium for members of this clade given the exceptionally greater diversity. *Colossendeis* has been reported from bathypelagic samples (Staples, 2007 and references therein) and a distribution with deep-sea currents and not the Antarctic Circumpolar Current are conceivable. It should be noted that whereas haplotypes were shared at least partly for *C. megalonyx* clade A between the two distant regions, no haplotype sharing was observed for *P. patagonica* clade ANT_D here. This indicates that, if at all, gene flow is extremely limited or represented a singular colonization event in *P. patagonica*.

Our enlarged sampling has extended the previously reported distribution ranges of some clades. *P. yepayekae* was found in the Strait of Magellan, which extends the occurrence of this species southwards (Weis et al., 2014). With the first records of specimens of clades ANT_F and ANT_C in the Eastern Weddell Sea, we could extend the distribution range of clades previously only reported from the Antarctic Peninsula (Harder et al., 2016). This is also the case for clade ANT_D where individuals from South Georgia were added to a clade previously only reported from the Antarctic Peninsula. It has been stated that narrow rather than broad distribution ranges might be the rule rather than the exception for sea spiders. Krabbe et al. (2010) postulated that most *C. megalonyx* clades have a narrow and allopatric distribution. However, analyzing more samples lead to the result of clades with a circumpolar distribution with isolation by distance (Dietz et al., 2015b). Thus, for *P. patagonica* s.l. we may also expect much broader distribution ranges when further material especially from unsampled locations is going to be included.

Furthermore, geographic separation between populations within clades (regional pattern) seems likely as we find subclades in several clades that are geographically separated. GMYC analyses revealed a subclade within *P. yepayekae* consisting of four individuals from the same area. A stronger effect of geographic separation can be seen for the second reported subdivision of the bPTP/ABGD clade by GMYC. For ANT_D there is a separation between an island population (South Georgia) and one from the Antarctic shelf (i.e., Antarctic Peninsula). Although supported by nuclear data, SUB_4 and SUB_5 clustered together in the ABGD analysis. Here, SUB_4 is represented by individuals from the Falkland Islands only.

Isolation of Falkland Island populations from those found on the rest of the South American continental shelf has also been reported for the isopod *Serolis paradoxa* (Leese et al., 2008).

We also found three new clades in the Eastern Weddell Sea only. It thus might be that different clades have different dispersal capabilities, however, as many habitats around the Antarctic shelf (e.g., Davis Sea and Dumont d'Urville Sea) and Subantarctic islands (e.g., Kerguelen Plateau) have been scarcely sampled, we cannot exclude that distribution ranges are generally broader than currently reported.

Which Clade Represents *P. patagonica* *Sensu Stricto*

The type specimen of *P. patagonica* (Hoek, 1881) has been collected from the Atlantic opening of the Strait of Magellan. In the absence of material from the Strait of Magellan, Weis et al. (2014) already proposed the Falkland clade as *P. patagonica* *sensu stricto*. Adding new samples, the Falkland clade also included samples from the Strait of Magellan, however it should be mentioned that this clade can geographically and genetically be subdivided into two sub clades. Both, GMYC and bPTP divided the Falkland clade into SUB_4 and SUB_5 that respectively included either samples from the Falkland Islands or the Strait of Magellan. Likely, specimens here assigned to clade SUB_5 represent the closest relatives of *P. patagonica* *sensu stricto*. However, ABGD results for the mitochondrial data combined SUB_4 and SUB_5 into a single clade. This was also supported by ITS data. Further information about the sub clades and the assignment of the type specimen might be obtained by a morphological reinvestigation of the type material in comparison with the new material from the Strait of Magellan.

Divergence Dates

In the absence of calibrated rates, molecular clock estimates using rates from other taxa can only be regarded as a rough proxy. For cold environments it might be assumed that mutation rate is lower as compared to temperate and tropic regions ("slow-rate hypothesis"; Bargelloni et al., 1994). However, evidence for this is still ambivalent (Held, 2001). Specifically, as we are addressing very recent divergence times it can be assumed that divergence times may be systematically higher than the rates inferred from rather deep calibration points (see Ho et al., 2005). The rate used and the error bars should thus be regarded as a rough orientation helping to interpret the radiation of *P. patagonica*. Even when considering the huge error bars, it is obvious that the divergence of the Subantarctic and the Antarctic super-clades took likely place in the Miocene after the opening of the Drake Passage. This indicates a single colonization event after the onset of the Polar Front and the Antarctic Circumpolar Current. The direction of the colonization (out of or into the Antarctic) remains unsolved in view of the limited number of outgroups. Also, the radiation of the many Antarctic *P. patagonica* species very likely started in the late Pliocene and increased during the Pleistocene (last 2.5 myr). Such patterns have been reported before (e.g., Held, 2000; Page and Linse, 2002; Thornhill et al., 2008; Krabbe et al., 2010; Leese et al., 2010; Hemery et al., 2012; Dietz et al., 2015a,b) suggesting that over evolutionary time scales the Polar Front has not been

an impermeable barrier to gene flow in general, though we did not observe across Drake Passage exchange in more recent clades of *P. patagonica* s.l. However, in view of the above-mentioned limitations of molecular clock calculations, we advise to use the divergence estimates made here with caution.

Comparing Species Complexes of *P. patagonica* and *C. megalonyx*

It becomes obvious that total divergence contained in *P. patagonica* s.l. exceeds the genetic divergence found within *C. megalonyx* (17 vs. 11% maximum pairwise distances, respectively). This suggests that species delimitation using morphological characters is more advanced in *Colossendeis* as compared to *Pallenopsis*. The description of *P. yepayekae* within *P. patagonica* s.l. shows that morphologically clearly distinguishable species exist, but no similarly detailed morphological inspections as within *Colossendeis* have been performed yet (Hodgson, 1907, 1908; Fry and Hedgpeth, 1969; Pushkin, 1993; Child, 1995; Dietz et al., 2013, 2015a). Thus, when comparing the complexes of *C. megalonyx* and *P. patagonica* side by side a similar proportion of the tree should be taken into consideration. When comparing pairwise COI sequence divergence between *C. megalonyx* (Krabbe et al., 2010; Dietz et al., 2015b) with all representatives of the Antarctic clade within *P. patagonica* s.l. a striking result is that barcode gap patterns look almost identical (Figure 3). Also, neither of the two species complexes shows a distinct barcode gap, pairwise sequence distances in the range of 2–5% are found at low frequencies. This is the reason for the more ambiguous ABGD results. However, the majority of inter-clade comparisons for both complexes are in the range of 6.5–9.5% (see Figure 3). These are values typically reported as interspecific.

Still, there is a substantial difference between this study and the study on *C. megalonyx* by Dietz et al. (2015b) in that all mitochondrial clades in *P. patagonica* s.l. (with the exception of ANT_L and ANT_H that are identical for ITS) are also supported by diagnostic ITS substitutions (Supporting Information Table S2), whereas in *C. megalonyx* there is strong evidence for hybridization across several clades that have even more than 7% COI divergence. The results for *P. patagonica* thus are similar to most other studies on Southern Ocean biota finding mitonuclear agreement (e.g., Leese and Held, 2008; Dietz et al., 2015a) and thus support of distinct species. It thus remains subject to discussion whether hybridization among *C. megalonyx* clade members as opposed to *P. patagonica* is possible due to the slower build-up of pre- or post-zygotic reproductive barriers.

One result of the direct comparison of both species complexes with the same molecular markers made here is that similar processes may have led to the divergence of distinct mitochondrial lineages. Assuming similar molecular clock rates, both have likely taken place in the same period (Pliocene/Pleistocene) characterized by drastic environmental changes between glacial and interglacial periods (see Thatje et al., 2005; Allcock and Strugnell, 2012). Yet, whereas for *C. megalonyx* hybridization of many of the species has been detected, this is not the case for *P. patagonica* s.l. Given the limited information on

the biology of the species reasons for this difference are difficult to estimate. One reason might be differences in the reproductive mode, another one differences in dispersal capabilities. Due to a lack of knowledge about reproduction within different sea spider species a direct comparison is not possible. However, larval stages and egg carrying males have never been reported for the genus *Colossendeis* possibly indicating low reproduction rate, whereas for *Pallenopsis*, males carry the eggs until hatching (benthic brooding) indicating a higher reproduction rate than in *Colossendeis*. Therefore, a low dispersal capability is assumed for *Pallenopsis* (except for occasional dispersal of adults, see below) while the situation in *Colossendeis* is unclear. The data for *C. megalonyx* Clade A (shared haplotypes between South Georgia and the Antarctic Peninsula) as well as most of the circumpolar clades in comparison to the many clades in *P. patagonica* that show rather narrow distribution ranges add further support for this difference in mobility. A lack of dispersal between isolated habitats can in principle promote the rise of reproductive barriers. Other possibilities include e.g., much stronger patterns of sexual selection and thus pre-zygotic mechanisms leading to faster complete lineage sorting. Also, *C. megalonyx* might show generally greater effective population sizes that counteract speciation. In view of similar divergence and diversity patterns for the COI data this seems, however, implausible. Perhaps the Antarctic Peninsula was colonized by active (i.e., walking) migration of *C. megalonyx* Clade A individuals from South Georgia through the deep sea after the end of the last glacial period. *Pallenopsis* has only been reported for the meso-pelagial but also in upper water plankton samples and drifting on jellyfish (Pages et al., 2007 and references therein). However, most likely the Antarctic Circumpolar Current prevents a drift across of the Antarctic Polar Front.

While for *C. megalonyx* we see strong evidence for an *in situ* evolution in Antarctica and migration to the Subantarctic (Clade B, M), this can neither be proved nor rejected for *P. patagonica* yet.

CONCLUSIONS

The results of our study on the sea spider *P. patagonica* support some, but not all of our initial hypotheses: (1) We find an increase of mitochondrial clades and an extension of distribution ranges with additional sampling. (2) Adding for the first time nuclear ITS data to verify the detected mitochondrial lineages in general found good agreement between both marker systems, i.e., no mitonuclear discordances. This is in disagreement to a recent report in *C. megalonyx* where strong evidence for hybridization and introgression was reported. Therefore, we suggest that the number of mitochondrial clades likely resembles the number of distinct species. However, application of state of the art species delimitation methods and analysis of both mitochondrial and nuclear genes does not lead to an unequivocal species delineation. Hence, future work needs to include more sets of characters for integrative taxonomy. The application of a molecular clock approach suggests that drivers of the biodiversity pump (speciation drivers) have acted at the same time scales

producing mainly young divergences in both *P. patagonica* s.l. and *C. megalonyx* but led to the formation of new species more efficiently in *P. patagonica* s.l.

AUTHOR CONTRIBUTIONS

Conceived the study: JSD, FL, and RRM. Laboratory analyses: JSD, AMH, and ARM. Bioinformatic analyses: JSD, FL. Wrote the paper: JSD, FL, and RRM.

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SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: <http://journal.frontiersin.org/article/10.3389/fevo.2016.00139/full#supplementary-material>

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Conflict of Interest Statement: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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